

7th October 2024



QIVIVE AND VARIABILITY DISTRIBUTIONS IN ELIMINATION USING TKPLATE

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Senior Scientific Officers

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OUTLINE

New Approach Methodologies and EFSA case studies

Human variability in metabolism and pathway-related uncertainty factors

TKPlate and QIVIVE

Moving Forward

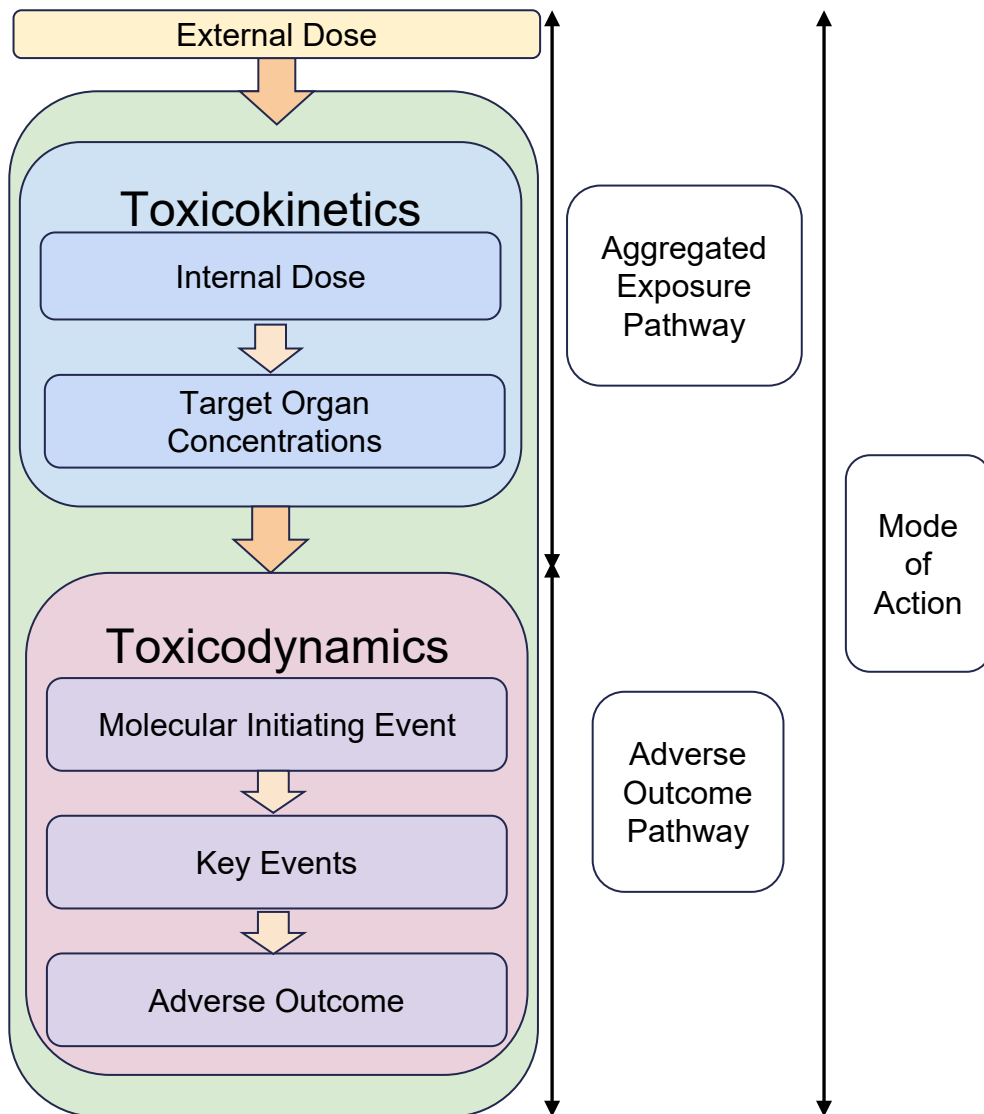




New Approach Methodologies and EFSA Case studies



UNDERSTANDING TOXICOKINETIC AND TOXICODYNAMIC PROCESSES IN CHEMICAL RISK ASSESSMENT



SCIENTIFIC REPORT OF EFSA

Modern methodologies and tools for human hazard assessment of chemicals¹

European Food Safety Authority^{2,3}

European Food Safety Authority (EFSA), Parma, Italy

*This scientific output, published on 11 July 2014, replaces the earlier version published on 24 April 2014**

ABSTRACT

This scientific report provides a review of modern methodologies and tools to depict toxicokinetic and toxicodynamic processes and their application for the human hazard assessment of chemicals. The application of these methods is illustrated with examples drawn from the literature and international efforts in the field. First, the concepts of mode of action/adverse outcome pathway are discussed together with their associated terminology and recent international developments dealing with human hazard assessment of chemicals. Then modern methodologies and tools are presented including *in vitro* systems, physiologically-based models, *in silico* tools and OMICs technologies at the level of DNA/RNA (transcriptomics), proteins (proteomics) and the whole metabolome (metabolomics). Future perspectives for the potential applications of these modern methodologies and tools in the context of prioritisation of chemicals, integrated test strategies and the future of risk assessment are discussed. The report concludes with recommendations for future work and research formulated from consultations of EFSA staff, expert Panels and other international organisations.

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KEY WORDS

mode of action, adverse outcome pathway, integrated testing strategy, physiologically-based models, *in silico*, OMICs

GUIDANCE

ADOPTED: 17 November 2021

doi: 10.2903/j.efsa.2021.7033

Guidance Document on Scientific criteria for grouping chemicals into assessment groups for human risk assessment of combined exposure to multiple chemicals

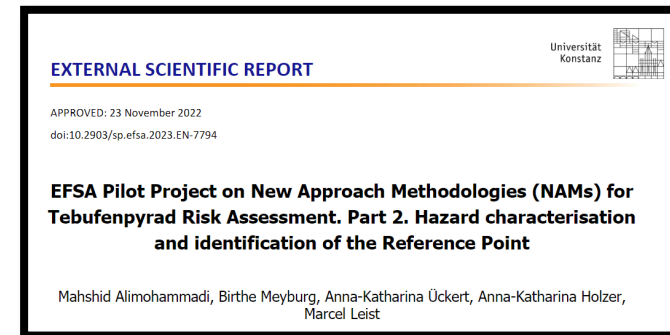
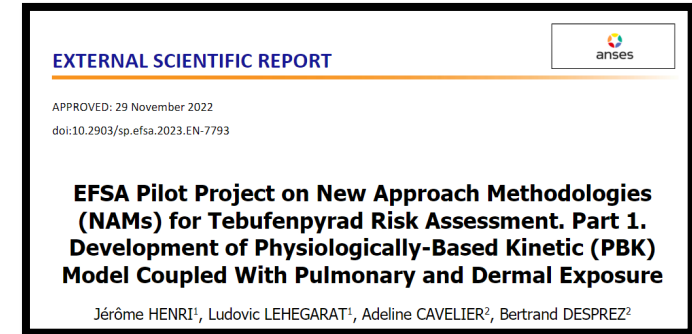
EFSA Scientific Committee,
 Simon John More, Vasileios Bampidis, Diane Benford, Claude Bragard,
 Antonio Hernandez-Jerez, Susanne Hougaard Bennekou, Thorhallur Ingi Halldorsson,
 Konstantinos Panagiotis Koutsoumanis, Claude Lambre, Kyriaki Machera, Hanspeter Naegeli,
 Soren Saxmose Nielsen, Josef Rudolf Schlatter, Dieter Schrenk, Vittorio Silano,
 Dominique Turck, Maged Younes, Emilio Benfenati, Amélie Crépet, Jan Dirk Te Biesebeek,
 Emanuela Testai, Bruno Dujardin, Jean Lou CM Dorne and Christer Hogstrand



EFSA NAMS CASE STUDY ON TEBUFENPYRAD

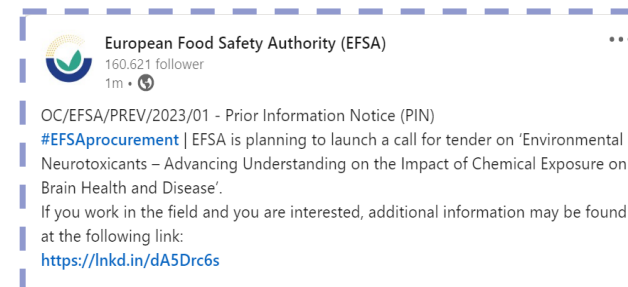
Aim: explore the use of NAMs to investigate the neurotoxicity potential of the pesticide Tebufenpyrad

- Part 1: 'Development of physiologically-based kinetic (PBK) model coupled with pulmonary and dermal exposure'
- Part 2: 'Hazard characterisation and identification of the Reference Point'



Q3 2020 – Q3 2022

A Workshop on the results of the project and follow up activities was organised in September 2022



EFSA NAMS CASE STUDY ON PFAS

External Scientific Report



APPROVED: 2 July 2024
doi: 10.2903/sp.efsa.2024.EN-8926

EFSA Project on the use of NAMs to explore the immunotoxicity of PFAS

Emanuela Corsini¹, Martina Iulini¹, Valentina Galbiati¹, Ambra Maddalon¹, Francesco Pappalardo², Giulia Russo², Ron L.A.P. Hoogenboom³, Karsten Beekmann³, Aafke W.F. Janssen³, Jochem Louisse³, Styliani Fragki⁴, Alicia Paini⁴

¹Università degli Studi di Milano, Italy; ²Università degli Studi di Catania, Italy
³Wageningen Food Safety Research, The Netherlands; ⁴ESQlabs GmbH, Germany

To develop and implement a NAM-based IATA for exploring the mode of action (MoA) for the observed immunosuppression effects and for addressing immunotoxicity of PFAS other than PFOS and PFOA, including the assessment of a common MoA and potency differences.

APCRA
Case
Study



EFSA NAMS CASE STUDY ON ESSENTIAL OILS

External Scientific Report

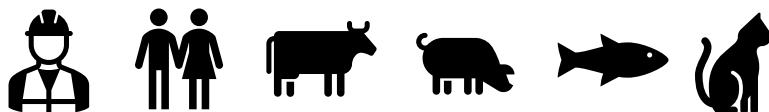


APPROVED: 15 May 2024
doi: 10.2903/sp.efsa.2024.EN-8820

EFSA Project on the use of NAMs
to explore interspecies metabolic differences on
essential oils as feed additives

Annelies Noorlander, Leonie Lautz, Wendy Jansen Holleboom,
Patrick P.J. Mulder, Geert Stoopen, Ans Punt

Wageningen Food Safety Research, the Netherlands



Design and conduct a series of NAM-based experimental studies for assessing qualitative and quantitative differences and similarities in metabolic competences across different target species for essential oils components and to conduct a quantitative in vivo in vitro extrapolation (QIVIVE) and comparison among species.

Table 5: Kinetic parameters for phase I metabolism of methyleugenol for the tested S9 fractions from different species

Species	Overall CL _{int} (μL/min/mg S9) ^a	1'-Hydroxylation (μL/min/mg S9) ^b	Detoxification (μL/min/mg S9)	% Bioactivation
Cat	17	5	12	29%
Chicken	26	2	24	8%
Cow	102	40	62	39%
Human	33	5	28	15%
Pig	69	27	42	39%
Rat	16	5	11	31%



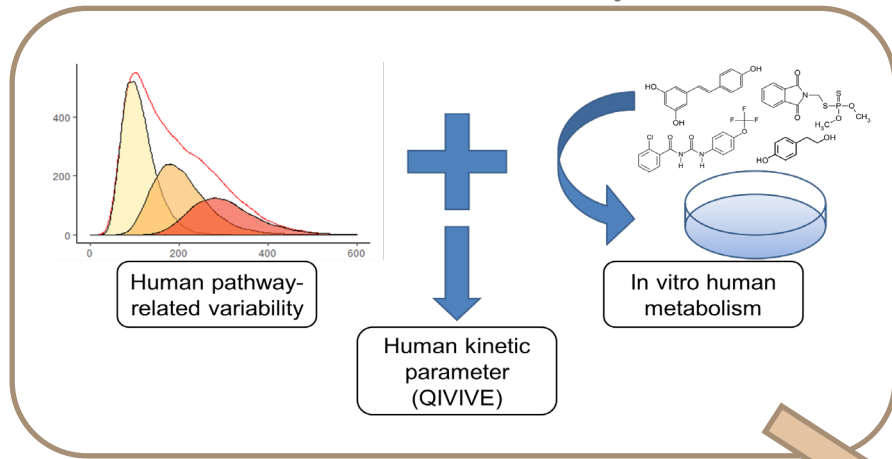


Human Variability in Metabolism and Pathway-related Uncertainty factors

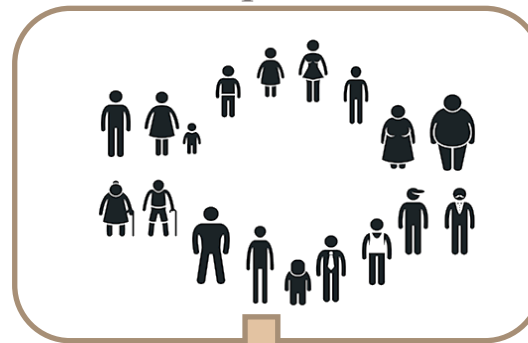


TK MODELLING AND APPLICATIONS IN HUMAN RISK ASSESSMENT

Kinetic Variability

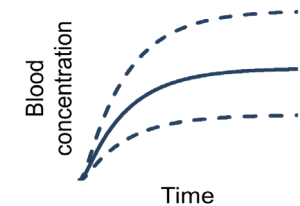


Population

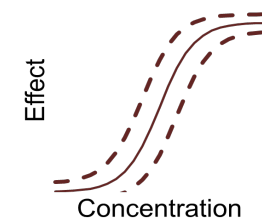


Risk Assessment

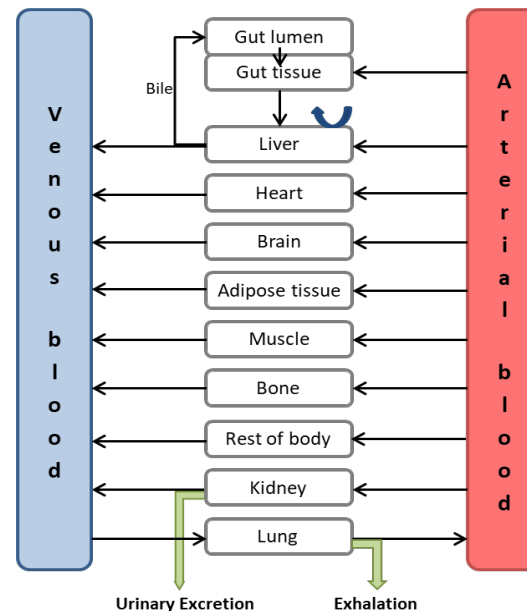
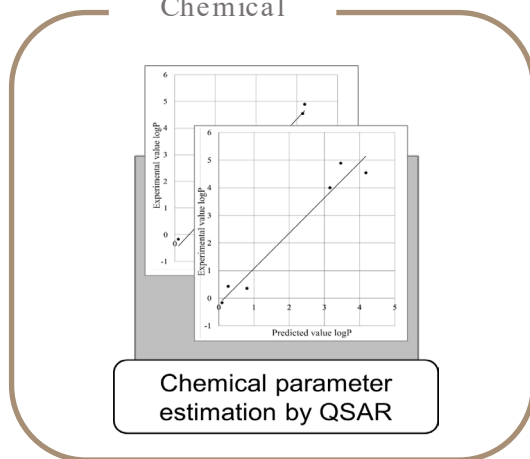
- Population specific simulations
- Integration of kinetic parameters
- Internal Dose estimation for target organs



- In vitro in vivo extrapolations for effects
- In vivo Dose-response modelling based on target organ concentrations



Chemical



QUANTITATIVE IN VITRO IN VIVO EXTRAPOLATION FOR HUMANS

- New data requirements for pesticides (283/284 2013) :
 - Scientific Opinion on use of comparative in vitro metabolism-Test Species vs Humans
 - Use human in vitro metabolism data and QIVIVE in RA
 - Case studies-prediction of human kinetics for food and feed chemicals (e.g. pesticides, contaminants)
 - Use of OHT 58 (TK) and OHT 201 (intermediate effects) to structure datasets
- Variability in metabolism within PBK models for QIVIVE



SCIENTIFIC OPINION

ADOPTED: 10 November 2021

doi: 10.2903/j.efsa.2021.6970

Scientific Opinion of the Scientific Panel on Plant Protection Products and their Residues (PPR Panel) on testing and interpretation of comparative *in vitro* metabolism studies

EFSA Panel on Plant Protection Products and their Residues (EFSA PPR Panel), Antonio F Hernandez-Jerez, Paulien Adriaanse, Annette Aldrich, Philippe Berry, Tamara Coja, Sabine Duquesne, Andreas Focks, Marina Marinovich, Maurice Millet, Olavi Pelkonen, Silvia Pieper, Aaldrik Tiktak, Christopher J Topping, Anneli Widenfalk, Martin Wilks, Gerrit Wolterink, Ursula Gundert-Remy, Jochem Louisse, Serge Rudaz, Emanuela Testai, Alfonso Lostia, Jean-Lou Dorne and Juan Manuel Parra Morte



EXTERNAL SCIENTIFIC REPORT

APPROVED: 22 February 2021

doi:10.2903/sp.efsa.2021.EN-6504

Modelling human variability in toxicokinetic and toxicodynamic processes using Bayesian meta-analysis, physiologically-based modelling and *in vitro* systems

Emanuela Testai, Camille Bechaux, Franca M. Buratti, Keyvin Darney, Emma Di Consiglio, Emma E.J. Kasteel, Nynke I. Kramer, Leonie S. Lautz, Nicoletta Santori, Zoi-Vasiliki Skaperda, Dimitrios Kouretas, Laura Turco, Susanna Vichi

OECD GUIDANCE ON THE USE OF PBK MODELS IN RA (2021)

Guidance document on the characterisation, validation and reporting of Physiologically Based Kinetic (PBK) models for regulatory purposes



Series on Testing and Assessment
No. 331



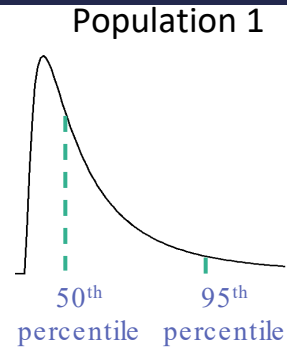
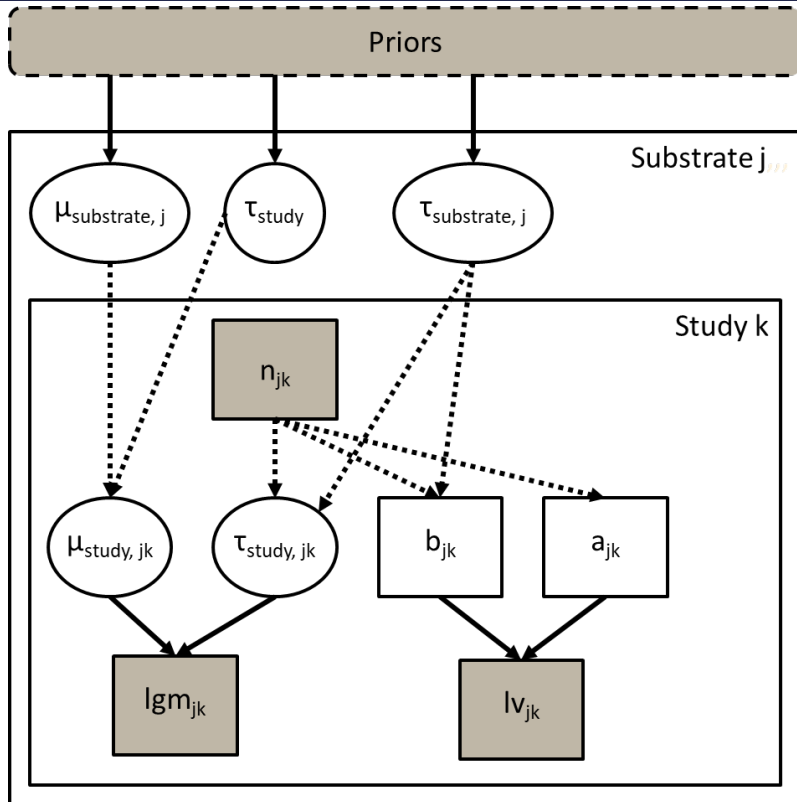
EFSA Case Studies on PBK Models in Annex

- Humans
- Fish
- Farm animals

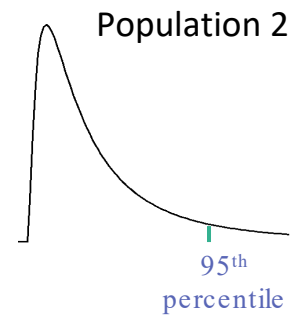
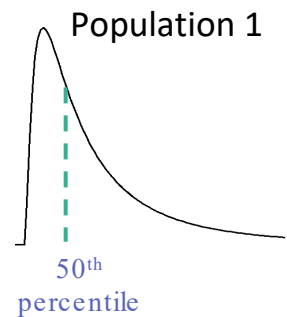
Chapter 2. PBK modelling workflow.....
2.1. Introduction
2.2. Step 1 – Scope and purpose of the model (problem formulation)
2.3. Step 2 – Model conceptualisation (model structure, mathematical representation)
2.4. Step 3 – Model parameterisation (parameter estimation and analysis).....
2.5. Step 3 – Computer implementation (Solving the equations)
2.6. Step 5 – Model performance
2.7. Step 6 – Model Documentation (reporting).....
2.8. Contextualisation of the PBK model for risk assessment.....
Notes.....
Chapter 3. Regulatory assessment of PBK models.....
3.1. Context and Implementation
3.2. Model validity
3.3. PBK Model Reporting Template
3.4. Checklist for Evaluation of Model Applicability



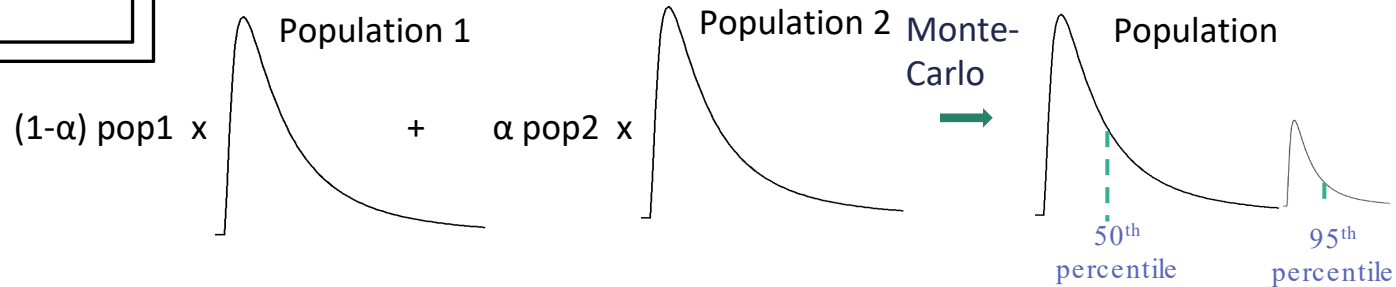
Probabilistic Uncertainty Factors



$$UF_{pop1} = \frac{P_{95}}{P_{50}}$$



$$UF_{pop2} = \frac{P_{95,pop2}}{P_{50,pop1}}$$



$$UF_{pop} = \frac{P_{95,pop}}{P_{50,pop}}$$

α : fraction of population 2 within general population



CYP3A4

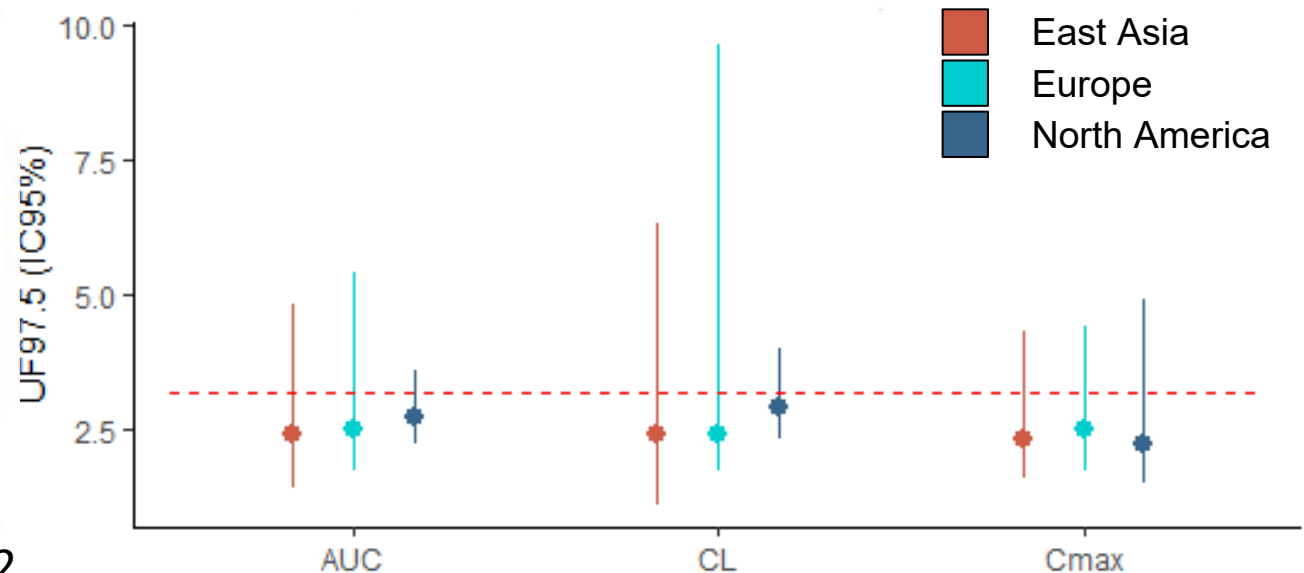
Variability and inter-individual differences in CYP3A4 metabolism:

ELS of TK data for 15 CYP3A4 probe substrates to collect parameters reflecting acute (C_{max}) and chronic exposure (clearance and AUC).

Variability after oral vs IV injection → 50% / 30% (AUC)

Healthy European, East Asian and North American adults showed generally similar CYP3A4-related UFs → limited interethnic differences.

	N substrate	N studies	n
Oral administration			
AUC (ng.h/ml/dose)	11	199	2921
Cl (ml/min/kg bw)	10	134	1603
C _{max} (ng/ml/dose)	12	221	3211
Intravenous administration			
AUC (ng.h/ml/dose)	4	40	577
Cl (ml/min/kg bw)	6	50	734



CYP3A4

These distributions allow to:

1. estimate UFs in the risk assessment process using variability distributions on metabolism,
2. Apply CYP3A4-related UFs in the risk assessment process for compounds for which *in vitro* CYP3A4 metabolism evidence are available,
3. Integrate CYP3A4-related variability distributions with *in vitro* metabolism data into physiologically based kinetic (PBK) models for quantitative *in vitro in vivo* extrapolation (QIVIVE).



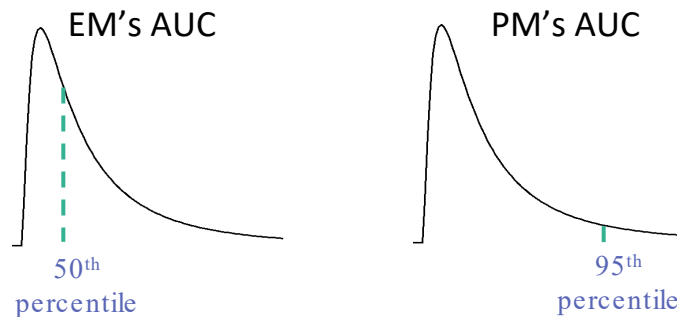
CYP2D6 POLYMORPHISM

How to derive UF that are protective for Poor Metabolisers ?

Caucasian mainly are Extensive Metabolisers and Poor Metabolisers (8%)

Variability in EM (AUC and clearance) of 50-100%

EM/PM AUC ratio range from 4 to 54



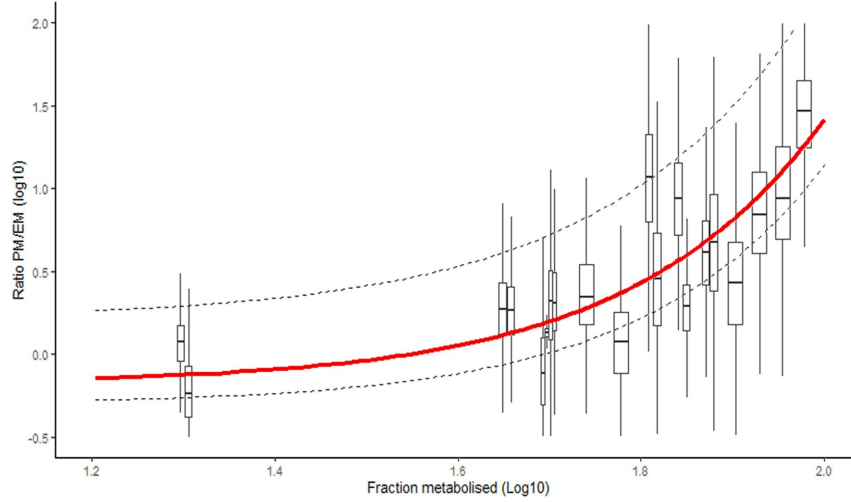
$$UF_{pop2} = \frac{P_{95,PM}}{P_{50,EM}}$$

PM UFs ranged for the AUC and clearance from 1.6-63.5 and 1.4-116 (P95)

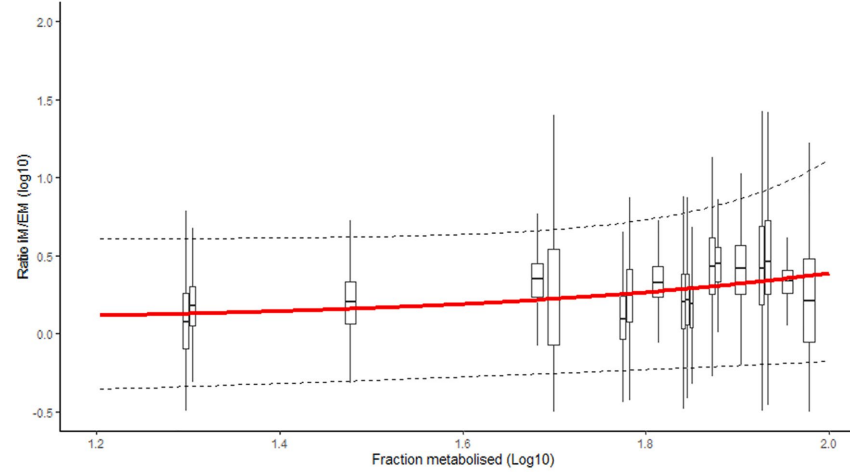


CYP2D6 polymorphism : Inter-phenotypic differences in Caucasian and Asian populations

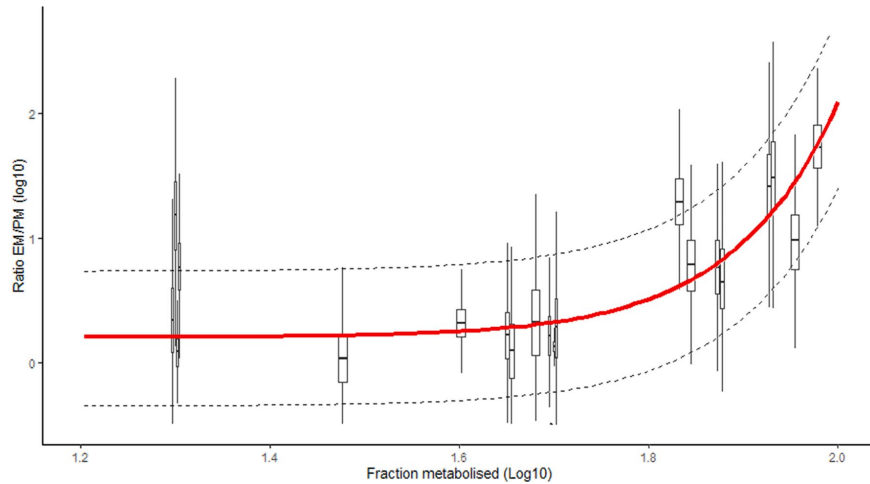
AUC Caucasian



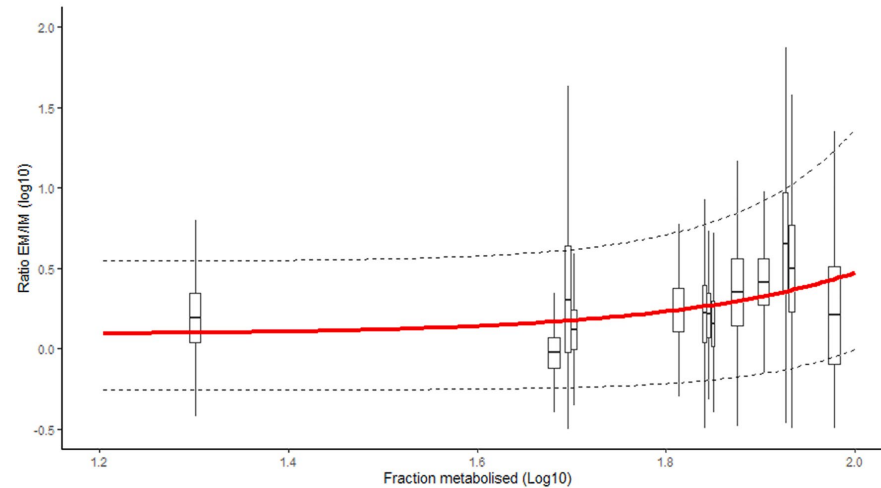
AUC Asian



Clearance Caucasian



Clearance Asian





TKPlate 1.0 and QIVIVE



TKPLATE 1.0: INFO PAGE AND GENERAL WORKFLOW

TKPlate - Default

https://r4eu.efsa.europa.eu/app/tktd

EFSA statistical models

Jean-Lou.DORNE@efsa.europa.eu Restart app Stop app Sign Out

v 1.0.20 - Manual - Report new issue

DOI: 10.5281/zenodo.7494936

TKPlate Interactive Modelling Platform

Info Input Forward Dosimetry Reverse Dosimetry Toxicodynamic DEB MixTox Report

WELCOME TO TKPLATE

This tool is EFSA's open source platform for the use of physiologically-based kinetic (PBK) models and New approach methodologies (NAMs) in human health, animal health and environmental risk assessment of chemicals. For more technical information, see:

- EFSA (European Food Safety Authority), Bossier H, Chau J, Cheikh N, Varewyck M, Verbeke T and Vergucht S. 2020 A Web-based open source tool for Toxicokinetic and Toxicodynamic modelling. EFSA supporting publication 2020:EN-1926. 25 pp doi:10.2903/sp.efsa.2020.EN-1926

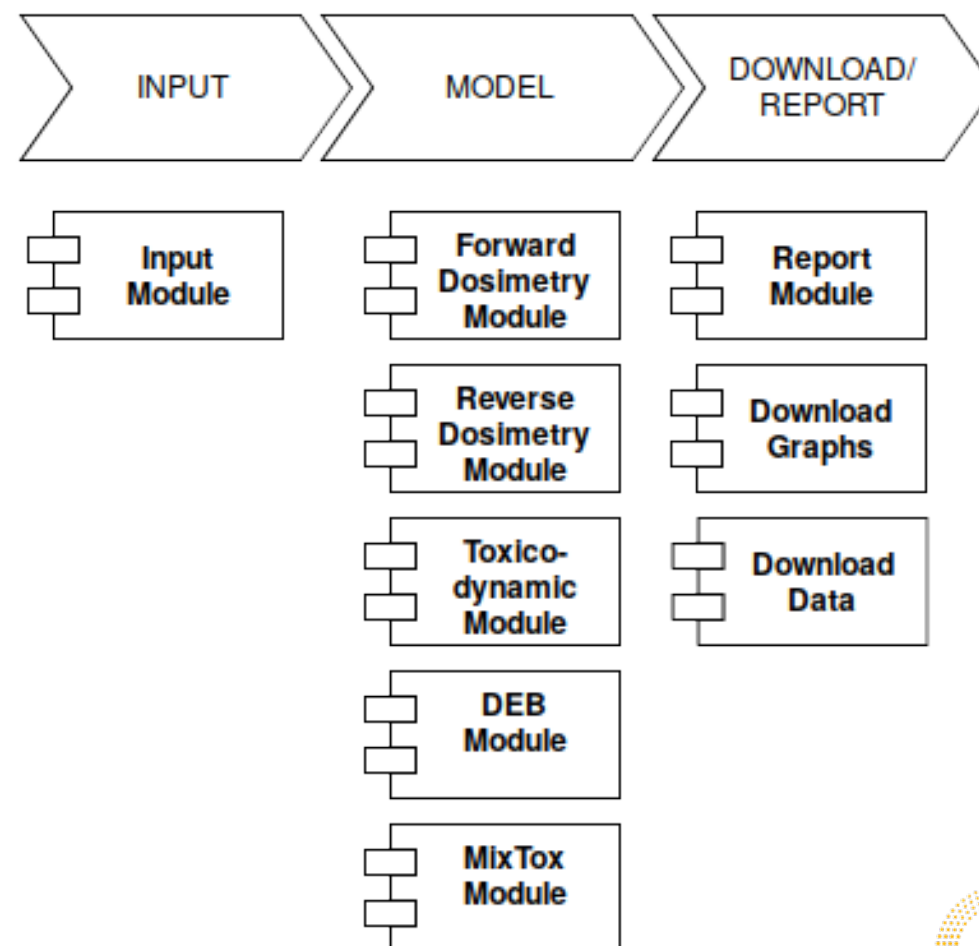
After 10 minutes of inactivity the app will stop working.

OVERVIEW

This info page is intended to give a quick overview of some of the possibilities within TKPlate. More details and information can be consulted in the [User Manual](#). The following sections describe the different modules that are available within the TKPlate 1.0 platform.

- The first module [Input] is the module where most input parameters must be provided. These are predominantly used for the forward dosimetry and benchmark dose modelling sections. Some parameter specific inputs such as biomonitoring data require uploading be uploaded in the modules requiring these data.
- The second module [Forward Dosimetry] is performing forward dosimetry where concentrations of chemicals in target organs of species are calculated after being exposed to an external dose. Variability is taken into account by means of Monte Carlo stochastic simulations. This module also provides the options to perform sensitivity analysis, check empirical data and compare different simulation results.
- The third module [Reverse Dosimetry] provides a means to perform reverse dosimetry which is used to predict a plausible exposure from bio-monitoring data.

TKPlate Workflow



TK PLATE 1.0 : PUBLICATION AND ASSOCIATED OUTPUTS

Further development of a web-based open-source platform for Toxicokinetic and Toxicodynamic modelling: TKPlate 1.0

Han Bossier, Joris Chau, Machteld Varewyck, Stephanie Vergucht and Tobias Verbeke

Open Analytics NV

Technical Report

APPROVED: 8 November 2023
doi: 10.2903/sp.efsa.2023.EN-8441



User Guide for TKPlate 1.0: An open access platform for implementing new approach methodologies in chemical risk assessment through toxicokinetic and toxicodynamic modelling

European Food Safety Authority (EFSA),
Han Bossier, José Cortiñas-Abrahantes, Keyvin Darney, Fotis Spyropoulos,
Leonie S Lautz, Pierre André Billat, Rémy Beaudouin, Florence Zeman, Cléo
Bodin, Jean Lou CM Dorne

Approved: 12 October 2023
DOI: 10.2903/j.efsa.2023.e211101

EDITORIAL

efsa JOURNAL

TKPlate 1.0: An Open-access platform for toxicokinetic and toxicodynamic modelling of chemicals to implement new approach methodologies in chemical risk assessment

Technical Report

APPROVED: 8 November 2023
doi: 10.2903/sp.efsa.2023.EN-8440

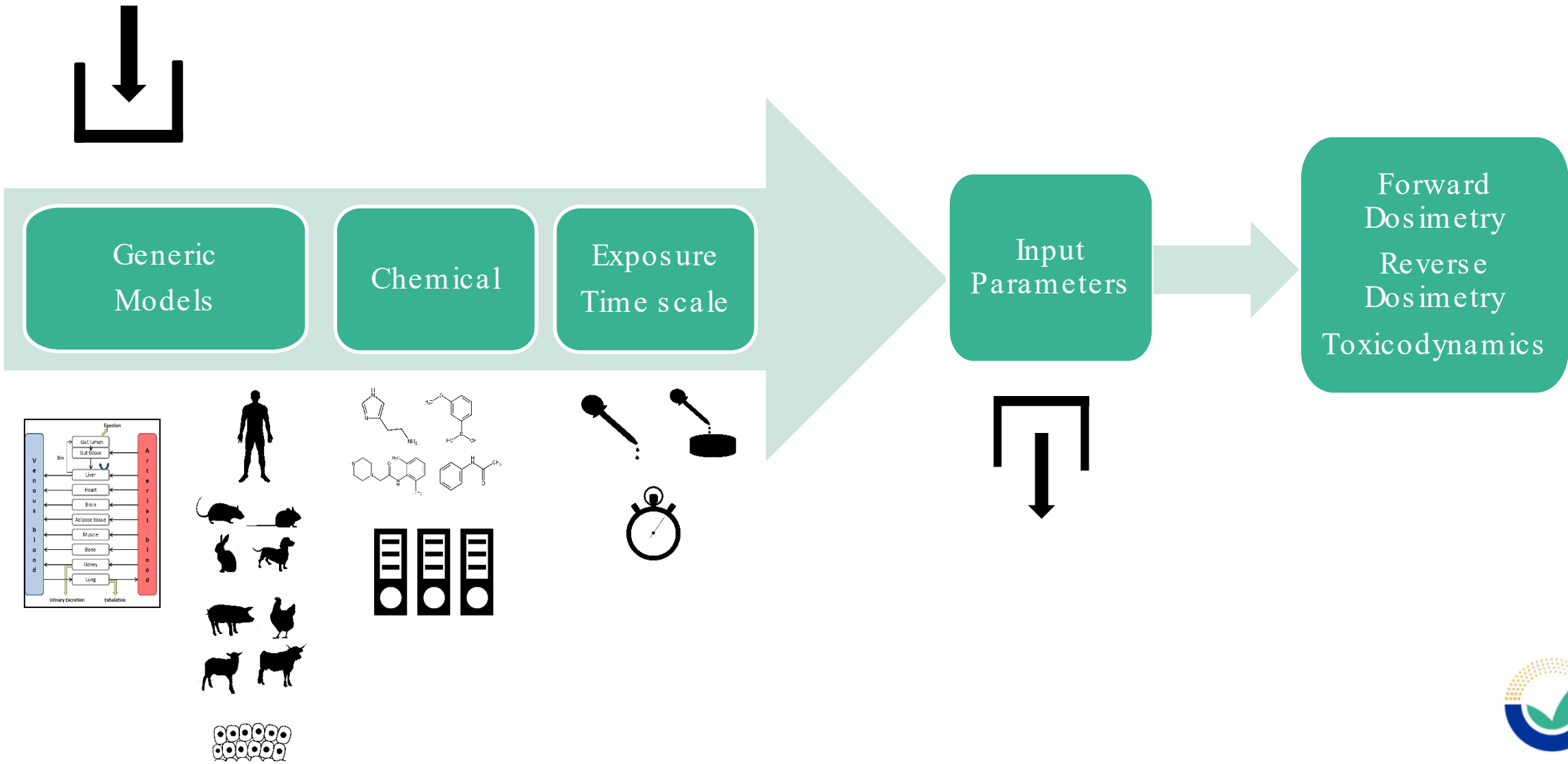


Applications of TKPlate 1.0 for toxicokinetic and toxicodynamic modelling of chemicals: Case Studies

European Food Safety Authority (EFSA),
Han Bossier, Fotis Spyropoulos, Keyvin Darney, Leonie S Lautz, Pierre André
Billat, Rémy Beaudouin, Florence Zeman, Cléo Bodin, José Cortiñas-
Abrahantes, Jean Lou CM Dorne



INPUT MODULE



INPUT MODULE : SELECTING MODEL, SPECIES AND CHEMICAL

Info | **Input** | Forward Dosimetry | Reverse Dosimetry | Toxicodynamic | DEB | MixTox | Report

Input

Model Type
Changing the model resets all filled-in parameter values.

generic physiologically based kinetic

Species
Changing the species resets all filled-in parameter values.

Human

Chemical

Available options for chemical selection

"(2s,3s)-n-[2-methoxy-5-(trifluoromethoxy)benzyl]-2-phenylpiperidin-3-amine"

User Defined Chemical

Show EPA CompTox Search URL

Parameters as Fixed Variables | **Parameters as Random Variables** | Initial State Values

Parameters allowed to take random values

Parameter name

BP

Transfer to 'Parameters as Random Variables' tab

Download and upload a parameter template

Download parameter template

Upload filled-in template

Browse... No file selected

Realistic Population

Use premade population

Calculate metabolism using:

Clearance (absolute) Clearance (relative) Km (mmol/L) & Vmax (mmol/min)

How to calculate the (relative) hepatic clearance?

Fill in own value in parameter table Use CV & CLi QIVIVE



INPUT MODULE : EXPOSURE SETTINGS, CHEMICAL PARAMETERS AND PHYSIOLOGICAL PARAMETERS

Exposure Settings ?

Dose unit

g mg microg

Time unit

d h min sec

Exposure Metric

absolute (mg) dose (mg/kg BW) rate (mg/kg BW / h)

Magnitude of each input (mg / kg BW / h)

Timepoint of first input (h)

Exposure time (h) (Cannot be zero)

Single or multiple doses?

Single

Multiple

Time Scale ?

Upload custom time points

time steps /h **Simulation duration (h)**

Chemical Specific Parameters

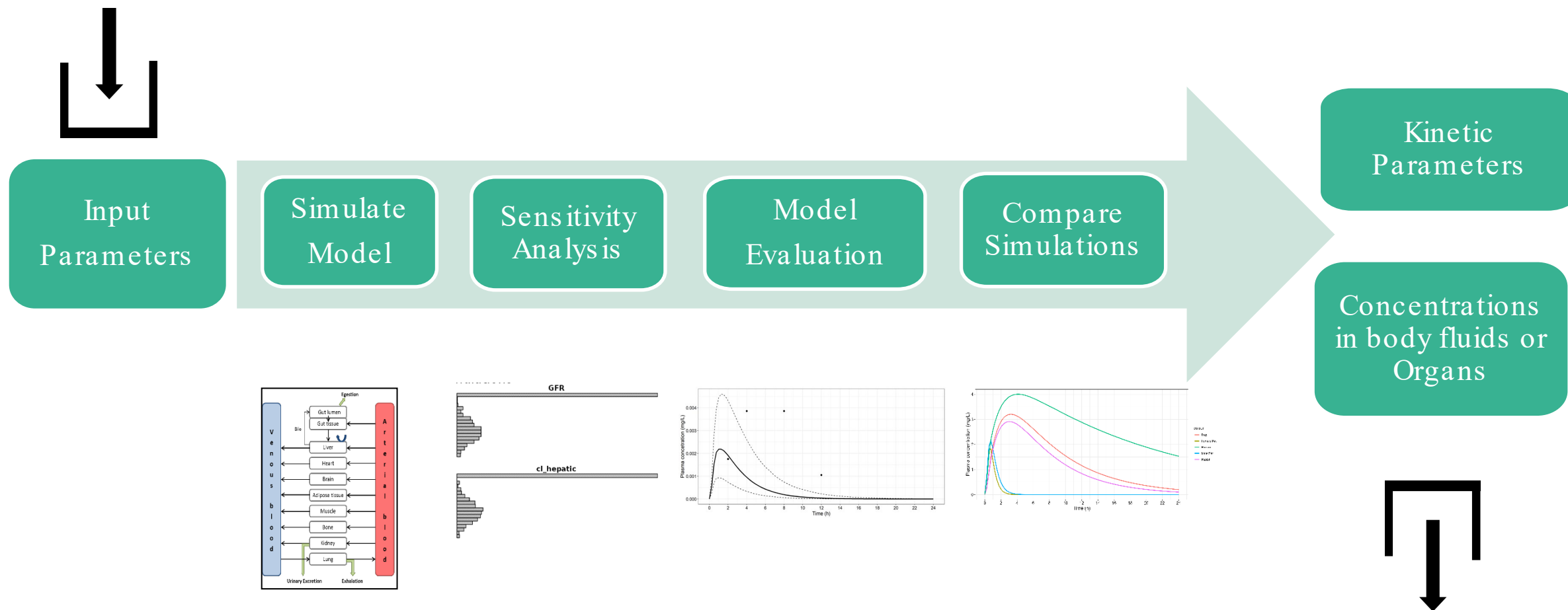
	Parameter	Value	Unit	Description
1	BP	1		Blood plasma ratio
2	cl_hepatic		L/min	Hepatic clearance
3	frac_abs_gut			Fractional absorption gut
4	FUP	1		Fraction unbound in plasma
5	GFR	0.06	L/min	Glomerular filtration rate
6	k_abs	0.01	min ⁻¹	Absorption rate constant (1st order)
7	MW	380.4	g/mol	Molecular Weight
8	P_adipose	1.29773		Blood tissue partition coefficient in the adipose
9	P_gut	1.321772		Blood tissue partition coefficient in the gut
10	P_kidney	2.109585		Blood tissue partition coefficient in the kidney
11	P_liver	2.579712		Blood tissue partition coefficient in the liver
12	P_rapid	2.664955		Blood tissue partition coefficient in the rapid compartment
13	P_slow	0.812618		Blood tissue partition coefficient in the slow compartment

Physiological Parameters

	Parameter	Value	Unit	Description
1	Q_adipose	0.42	L/min	Blood flow in adipose
2	Q_gut	1	L/min	Blood flow in gut
3	Q_kidney	1.23	L/min	Blood flow in kidney
4	Q_liver	0.41	L/min	Blood flow in liver
5	Q_rapid	1.28	L/min	Blood flow in rapid compartment
6	Q_slow	1.51	L/min	Blood flow in slow compartment
7	V_adipose	20.28	L	Volume of the adipose
8	V_gut	1.22	L	Volume of the gut
9	V_kidney	0.39	L	Volume of the kidney
10	V_liver	1.99	L	Volume of the liver
11	V_rapid	2.12	L	Volume in the rapid compartment
12	V_slow	36.74	L	Volume of the slow compartment



FORWARD DOSIMETRY : WHAT THE BODY DOES TO THE CHEMICAL (FROM OUTSIDE TO INSIDE)



Predicting what the body does to the chemical (Toxicokinetics):

- Absorption, metabolism, elimination : Kinetic parameters
- Residues in blood, urine and organs of interest (e.g. liver, kidney etc..)



FORWARD DOSIMETRY: OUTPUTS AND MODEL EVALUATION

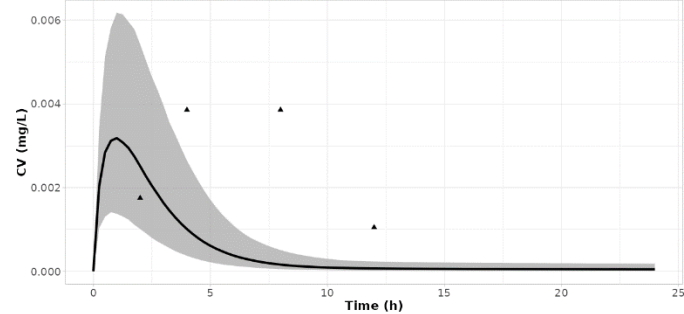
Outputs: Time Concentration Profiles/Kinetic Parameters

Simulations for single Species

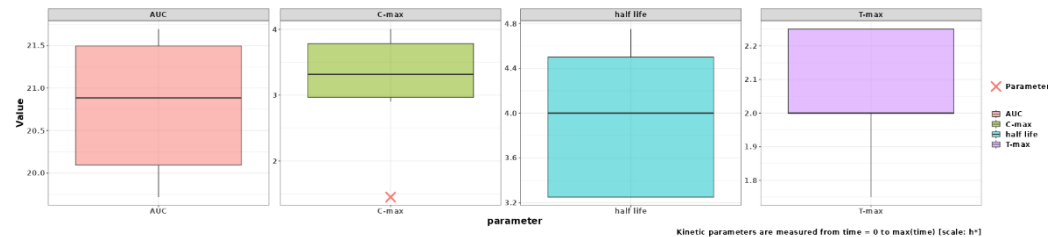
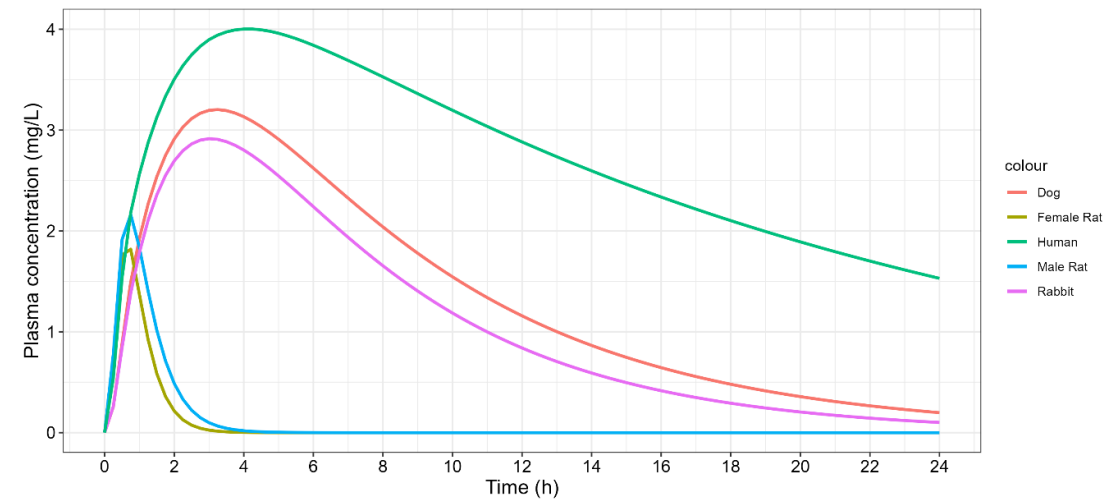
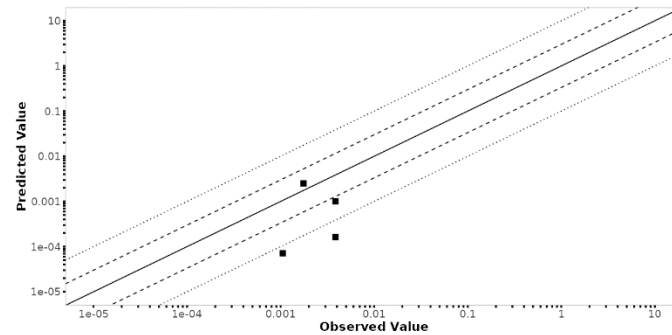
Compare Simulations for multiple species

MC simulated trajectory

Median, lower-2.5% and upper 97.5% quantiles, pointwise in time



Predicted vs observed concentrations.



Kinetic parameters are measured from time = 0 to max(time) [scale: h⁻¹]



TK PLATE QIVIVE MODULE

❖ CL_{int} unit $\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$

→ $\text{ml}/(\text{mg prot} \cdot \text{min})$

In vitro results

K_m & V_{max} → CL_i ($\text{pmol product}/\text{nmol P450} \cdot \text{min} \cdot \mu\text{M}$)

Relative Abundance ($\text{pmol P450}/\text{mg prot}$)

$MPPGL = 40 \text{ mg prot}/\text{g liver}$

$CL_{in vivo} = CL_i * RA * MPPGL / 1000$

enzyme	CV	CL_{int}	unit
2C8	0.3		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
2B6	0.3		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
3A4	0.5		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
2C9	0.5		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
2A6	0.3		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
2D6	0.5		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
1A1	0.3		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
1A2	0.35		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
2C19	0.5		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
2C18	0.3		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
3A5	0.3		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$
esterases	0.4		$\text{nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$



HANDLING POLYMORPHISM : A CASE STUDY

❖ Considering a compound metabolised by CYP3A4 and 2D6 in humans

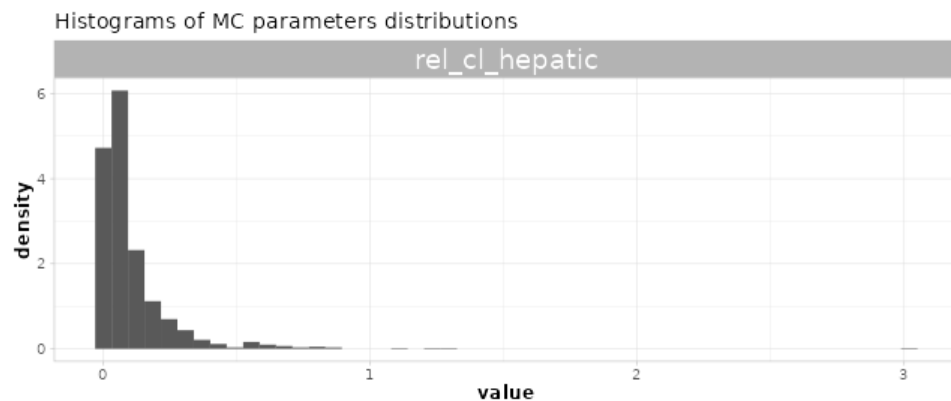
- $CL_{CYP3A4} = 0.001 \text{ nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$

❖ CYP2D6 Extensive metabolisers and poor metabolisers Ratio of 5 for Clint

- $CL_{int,CYP2D6} = 0.008 \text{ nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$

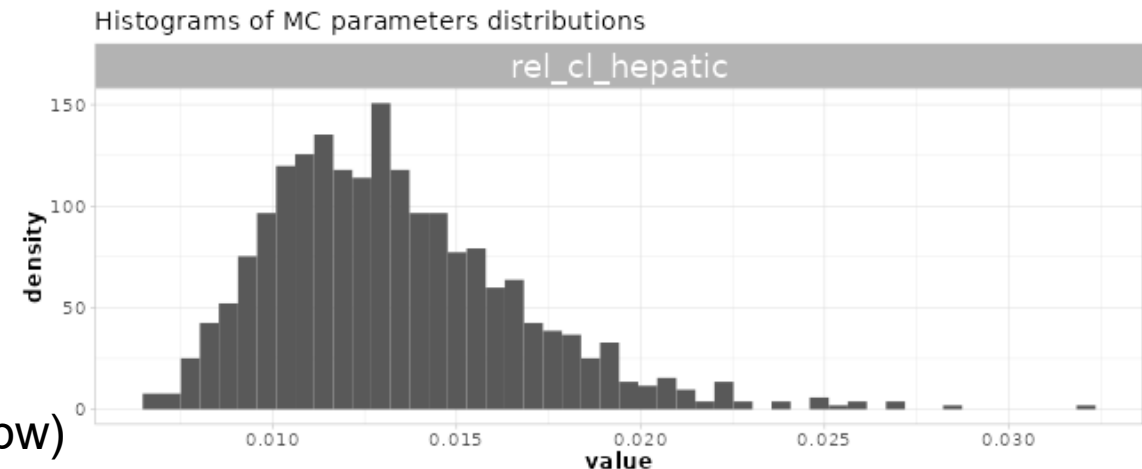
- $CL_{int,CYP2D6} = 0.04 \text{ nmol product}/(\text{mg prot} \cdot \text{min} \cdot \mu\text{M})$

PM



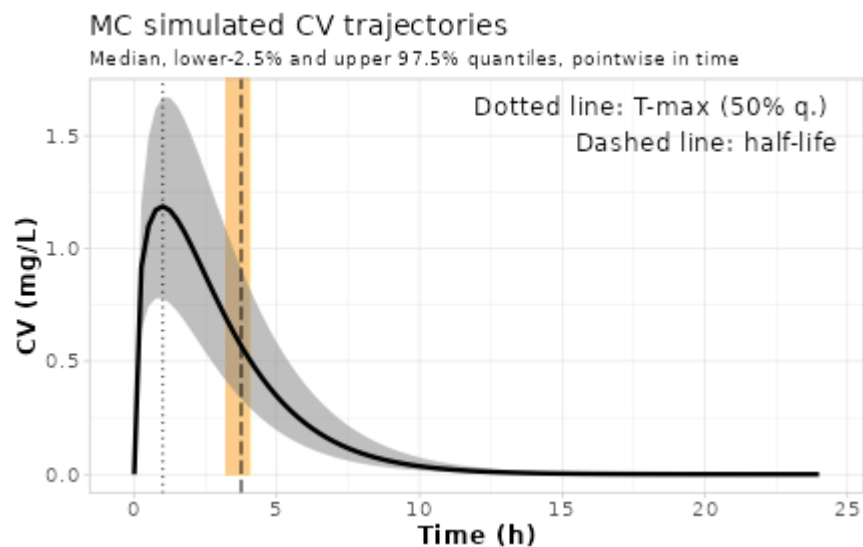
Estimated CL (L/min/kg bw)

EM

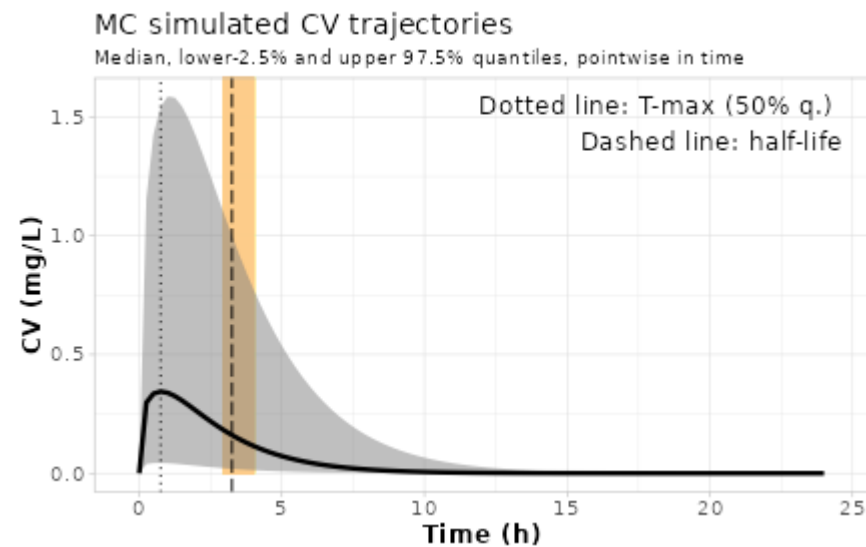


SIMULATING THE KINETIC PROFILES IN CYP2D6 EXTENSIVE AND POOR METABOLISERS

PM



EM



	0%-quantile	2.5%-quantile	50%-quantile	97.5%-quantile	100%-quantile
T-max (h)	7.50E-01	7.50E-01	1.00E+00	1.00E+00	1.25E+00
C-max (mg/L)	5.41E-01	7.78E-01	1.19E+00	1.67E+00	1.96E+00
Half-life (h)	2.50E+00	2.50E+00	2.75E+00	3.00E+00	3.25E+00
AUC (h * mg/L)	2.00E+00	3.01E+00	4.90E+00	7.54E+00	9.30E+00

	0%-quantile	2.5%-quantile	50%-quantile	97.5%-quantile	100%-quantile
T-max (h)	7.50E-01	7.50E-01	7.50E-01	1.00E+00	1.25E+00
C-max (mg/L)	9.76E-03	4.36E-02	3.44E-01	1.59E+00	2.40E+00
Half-life (h)	2.25E+00	2.25E+00	2.50E+00	3.00E+00	3.50E+00
AUC (h * mg/L)	3.31E-02	1.48E-01	1.23E+00	7.05E+00	1.25E+01



New Human models

- Refine QIVIVE models
- Generic models for a range of metabolic pathways
- Introducing a microbiome compartment
- Pregnancy/ Gestational models
- Ontogenesis of enzymes
- Generic models for metabolites (detoxification/bioactivation)
- Reverse dosimetry models for metabolites/HBM/urine etc
- Bringing a module for chemical-specific PBK models
- Bioaccumulation models (EFSA/ECHA)



MOVING FORWARD



- TKPlate Training

- Pilot planned for EFSA Staff and experts on 21-22 November 2024
- Pending Feedback second training with ECHA planned for Jan 2025
- Moving towards recorded online training available open access ?

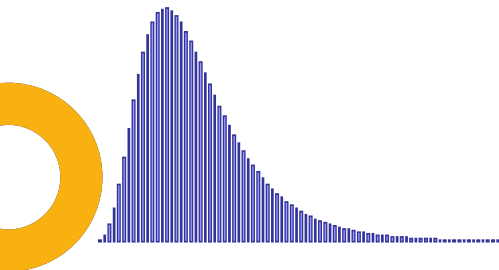


- Moving towards application of QIVIVE and TKPlate in EFSA panels

- Communicating outputs to panels through practical case studies
- Difference in NAM acceptance across panels : regulations, history
- Contaminants, pesticide, feed additive and novel food applications
- Change management may be needed: from in vivo to NAMs

- EC/International collaborations to implement models in RA

- PARC/ASPIS EU : follow up on models and data developed, uptake TKPlate
- Future APRCA case study on use of TKPlate for QIVIVE



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